

## Heterogeneous Clinical Characteristics of Allan-Herndon-Dudley Syndrome with SLC16A2 Mutations

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**Purpose:** The purpose of this study was to expand our understanding of phenotypic and genetic variation in Allan-Herndon-Dudley syndrome (AHDS), which is a rare X-linked mental retardation syndrome characterized by hypotonia, generalized spasticity, and moderate-to-severe psychomotor retardation. AHDS is caused by a mutation of solute carrier family 16 member 2 (*SLC16A2*), which encodes monocarboxylate transporter 8 (MCT8), a transporter of triiodothyronine (T<sub>3</sub>) into neurons.

**Methods:** We enrolled nine patients with AHDS from unrelated families, except for two patients who were cousins, through a retrospective chart review. Clinical features, brain imaging, electroencephalograms, thyroid hormone profiles, and genetic data were reviewed retrospectively and compared with previously reported cases.

**Results:** We found three novel and five previously reported pathogenic variants in nine patients from eight families. All patients presented with hypotonia, spasticity, severe developmental delay, and elevated serum T<sub>3</sub> levels. Cataplexy, which is a previously unreported phenotype, was found in two patients with the same mutation. In our cohort, seizures were uncommon (n=1) but intractable.

**Conclusion:** This study broadens the known phenotypic variations of AHDS, ranging from relatively mild global developmental delay to a severe form of encephalopathy with hypotonia, spasticity, and no acquisition of independent sitting. The syndromic classification or genetic etiology of global developmental delay is extremely heterogeneous; therefore, early clinical suspicion is challenging for clinicians. However, severe mental retardation with hypotonia, spasticity, and elevated serum T<sub>3</sub> levels in boys is a highly suspicious clinical clue for the early diagnosis of AHDS.

**Keywords:** Allan-Herndon-Dudley syndrome; SLC16A2 protein, human; Mental retardation, X-linked; Cataplexy

## Introduction

Allan-Herndon-Dudley syndrome (AHDS) is a rare disease characterized as an X-linked mental retardation syndrome that is associated with neuromuscular involvement and thyroid function abnormality. The first case of AHDS was reported by Allan et al. [1] in 1944, but its association with solute carrier family 16 member 2 (*SLC16A2*) mutation was first described in 2004 [2,3].

*SLC16A2* encodes monocarboxylate transporter 8 (MCT8), a transmembrane transporter of the active form of thyroid hormone, triiodothyronine ( $T_3$ ) [4]. Thyroid hormones are important for neurodevelopment and normal growth. Because MCT8 is highly expressed in the brain–blood barrier and neuronal cells of the developing brain [5], MCT8 defects result in severe psychomotor deficits including central hypotonia, generalized spasticity, dystonia, and moderate-to-severe intellectual disability [2,3]. Impaired myelination on brain magnetic resonance imaging (MRI) has been reported in the majority of affected patients, but the heterogeneity of MRI findings needs further evaluation [6,7].

The thyroid profile of patients with AHDS shows elevated  $T_3$  levels, low-to-normal free tetraiodothyronine ( $T_4$ ) levels, and normal-to-high thyroid-stimulating hormone (TSH) serum levels [2,3]. Due to the elevated serum levels of  $T_3$ , other organs, such as the heart, muscles, and liver, which are less dependent than the brain on the MCT8 protein for transporting thyroid hormones into the cell, are affected by peripheral thyrotoxicosis, the symptoms of which include muscle wasting, low body weight, and tachycardia [8,9].

The phenotypic spectrum of AHDS is highly varied, including patients who achieve independent walking with mild intellectual disability to those with severe encephalopathy [10,11]. The phenotype-genotype correlation is not yet understood fully, but residual protein activity is suggested to be associated with clinical severity [12].

Herein, we present nine patients who were diagnosed with genetically confirmed AHDS. Their clinical, laboratory, radiologic, and genetic characteristics were described in detail to broaden the scope of known phenotypic characteristics and to expand our understanding of the disease.

## Materials and Methods

### 1. Patients

Nine male patients aged from 0 to 16 years were diagnosed with AHDS at Seoul National University Children's Hospital from January 2010 to January 2020. The diagnoses were made by a clinical neurologist based on the patients' clinical presentation and thyroid

hormone profile followed by confirmative genetic testing. For the patients diagnosed with AHDS based on molecular analysis, we performed a detailed retrospective review of medical records including clinical characteristics, laboratory test results including the thyroid hormone profile, brain MRI, and electroencephalography (EEG).

### 2. Molecular analysis

Genomic DNA was extracted from peripheral blood from the patients and/or their family members by standard procedures after obtaining informed consent. Genetic variants were analyzed using targeted panels or whole-exome sequencing. The exon sequences and their flanking intronic sequences were identified through polymerase chain reaction using genomic DNA. The DNA sequences were mapped to and analyzed in comparison with the published human genome build (UCSC hg19 reference sequence [RefSeq]). The targeted coding exons and splice junctions of the known protein-coding RefSeq genes were assessed for the average depth of coverage and data quality threshold values. All reportable sequence variants below the threshold quality were confirmed by Sanger sequencing analysis using a separate DNA preparation. The DNA sequences were aligned to a human reference sequence (NCBI Accession No. NM\_006517.5 for *SLC16A2*).

*In silico* analysis of the missense variants was performed using the CADD, SIFT, and Polyphen-2 programs, which evaluate the predicted deleteriousness and functional effects of each variant. The PhyloP program (phylogenetic *P* values) was used to measure evolutionary conservation. The minor allele frequency of variants was determined using data from gnomAD.

This study was approved by the Institutional Review Board (IRB) of the Seoul National University Hospital (IRB No. 2101-173-1191). The requirement to obtain written informed consent from the patients was waived due to the retrospective nature of our study.

## Results

### 1. Demographics and clinical presentation of AHDS syndrome

All nine patients were male and born from nonconsanguineous parents of Korean ethnicity. Table 1 summarizes the clinical manifestations and examination results of the nine patients who were genetically confirmed to have AHDS.

### 2. *SCL16A2* mutation analysis and family history

We found three novel mutations (1396G > C [p.A466P], c.80delG [p.G27Vfs\*57], c.58C > T [p.Q20X]), and five previously report-

**Table 1.** Phenotypic characteristics and genotypes of the nine patients with Allan-Herndon-Dudley syndrome

Characteristic	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8	Patient 9
Gender	M	M	M	M	M	M	M	M	M
Onset age	6 months	3 months	5 months	6 months	6 months	6 months	4 months	12 months	11 months
Age at last visit	3 years	4 years	2 years	4 years	7 years	2 years	3 years	8 years	5 years
Perinatal features									
Prematurity	-	-	-	-	-	-	-	-	-
Low birth weight (<2.5 kg)	+	-	-	-	-	-	-	-	-
Perinatal problems	+	+	-	-	+	-	-	+	-
	Neonatal jaundice	Respiratory distress			Weak fetal movements			Respiratory distress	Neonatal jaundice
Neurologic features									
Hypotonia	+	+	+	+	+	+	+	+	+
Language	No words	No words	No words	No words	No words	No words	No words	Few words	No words
Motor performance	Non-ambulatory	Non-ambulatory	Non-ambulatory	Non-ambulatory	Non-ambulatory	Non-ambulatory	Non-ambulatory	Obedied simple commands	Obedied simple commands
	Incomplete head control	No head control	No head control	No head control	No head control	Head control incomplete; rolled over	Head control incomplete	Ambulatory	Ambulatory
							Sat with support	Independent ataxic gait	Walked with holding
Spasticity/hyperreflexia	+/+	+/+	+/+	+/+	+/+	+/+	+/+	+/+	+/+
Dystonia/choreoathetosis	-/-	+/-	-/-	+/-	+/-	+/-	+/-	+/-	+/-
Myoclonus/startles	+	-	+	+	-	+	+	+	-
Epilepsy	-	-	-	-	+	-	-	-	-
Cataplexy	-	-	-	-	-	-	-	+	+
Brain MRI abnormality	+	+	+	+	+	+	+	-	-
Delayed myelination	+	+	+	-	UC	+	+	-	-
Thin corpus callosum	-	+	-	+	UC	+	+	-	-
Cerebral atrophy	-	+	-	-	UC	+	+	-	-
EEG	UC	Intermittent slow wave	Normal	Intermittent slow wave	Focal spike discharges	Normal	UC	UC	UC
Growth parameters									
Microcephaly (<3 p)	-	-	-	-	+	-	-	-	-
Short stature (<3 p)	-	-	-	-	-	-	UC	-	-
Low weight (<3 p)	-	+	-	+	+	-	UC	-	-

(Continued to the next page)

**Table 1.** Continued

Characteristic	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8	Patient 9
Dysthyroidism	+	+	+	+	+	+	+	+	+
Skeletal abnormality	-	+	-	-	+	-	+	+	-
		Hip dislocation			Bilateral coxa valga, macrodactyly, Flexion deformity in left toes		Hip dislocation	Pes planovalgus	
Ocular abnormality	-	-	UC	UC	UC	+	-	-	-
						Strabismus			
Other		Hyperammonemia			Cryptorchidism			Patent ductus arteriosus	
Elevation of lactic acid	-	+	+	-	+		+	+	-
		Lactic acidosis	Lactic acidosis		Mild elevation		Mild elevation	Mild elevation	
Genomic variant NCBI Accession No. NM_006517.5	Exon 1 deletion	c.80delG	c.1026+1G>A	c.439G>A	c.449C>T	c.58C>T	Exon 5 deletion	c.1396G>C	c.1396G>C
Protein change	-	p.G27Vfs*57	-	p.G147R	p.A150V	p.Q20X	-	p.A466P	p.A466P
<i>In silico</i> analysis									
CADD score				29.4	24.2			28.1	28.1
SIFT									
Score				0.02	0.01			0	0
Prediction				Deleterious	Deleterious			Deleterious	Deleterious
Polyphen-2									
Score prediction				0.996	0.637			1.000	1.000
				Probably damaging	Possibly damaging			Probably damaging	Probably damaging
PhyloP score (mammalian)				2.336	2.336			2.267	2.267
Minor allele frequency		0.0	0.0	0.0	0.0	0.0		0.0	0.0
Mode of inheritance	Mat	Mat	UC	Mat	<i>De novo</i>	<i>De novo</i>	UC	Mat	Mat

-, negative finding; +, positive finding; MRI, magnetic resonance imaging; UC, unchecked; EEG, electroencephalography; NCBI, National Center for Biotechnology Information; Mat, maternal inherited.

ed mutations (c.449C > T [p.A150V], c.439G > A [p.G147R], c.1026+1G > A, exon 1 deletion, exon 5 deletion) in nine patients from eight families. Five patients inherited the mutation from their mother and two patients had *de novo* mutations. In two patients, maternal carrier status was not checked. The mutations in the *SLC16A2* are summarized in Table 1 and Fig. 1.

Family histories were reviewed and relatives with cerebral palsy of unknown etiology were observed in three patients from two families. Patients 8 and 9 were maternal cousins and patient 3 had a maternal uncle bedridden with spastic paraplegia, who was considered to have cerebral palsy from an unknown cause.

### 3. Perinatal problem

Antenatal problems were reported in one of the nine patients. Weak fetal movement, a small biparietal diameter, and low body weight were observed in patient 5, but delivery was uneventful. All patients were born full-term. Neonatal problems were present in three patients (33.3%). The patients with neonatal jaundice (n = 2) were treated by phototherapy, those with transient respiratory distress (n = 2) did not need ventilator support, and one patient with patent ductus arteriosus (n = 1) required a ligation operation. Patient 8 presented with neonatal jaundice, transient respiratory distress, and patent ductus arteriosus; however, patient 9, who had the same mutational variant as patient 8, did not experience any of these problems.

### 4. Growth

The growth parameters at birth were all normal for all patients except patient 1, who had low birth weight. Although all patients except one had a normal body weight at birth, poor weight gain was present in three patients. Microcephaly was present in patient 5, who also had low body weight (below the first percentile). Short stature was not observed in any of the nine patients during the follow-up period.

### 5. Neurologic symptoms

All patients visited the clinic for evaluations of motor developmen-

tal delay. The median age at the first visit was 6 months (range, 3 to 12) and the median age at the last follow-up was 4 years (range, 2 to 8). In seven patients, the initial symptom was an inability to control the head at the age of 3 to 6 months. In patients 8 and 9, who were maternal cousins with the same variant, developmental delay was noticed at 11 to 12 months of age because they could not sit alone by then. These two patients could walk with a dystonic and ataxic gait at 4 years old. The other seven patients who could not walk independently showed various patterns of motor development during the follow-up period, as follows: no head control (n = 2), partial head control (n = 3), partial head control and rolling over (n = 1), and partial head control and sitting with assistance (n = 1). Axial hypotonia was observed in all patients (n = 9). Pyramidal symptoms were also present in all patients, including spasticity (n = 9) and hyperreflexia (n = 9). Extrapyramidal symptoms were also frequent, including dystonia (n = 7) and choreoathetosis (n = 1). Notably, patient 9 showed no definite abnormality on the initial neurologic examination at the age of 11 months, and did not demonstrate axial hypotonia at that time. Other pyramidal and extrapyramidal features became evident and progressed over time.

Startles or myoclonic movements were common symptoms (n = 5). This symptom was triggered especially when the patients were about to sleep or were stimulated by sounds. Four patients underwent an EEG for startles or myoclonic movements to differentiate them from seizures. The EEG records from two patients were normal and the other two patients showed intermittent high-amplitude delta activities without definite epileptic discharges.

Seizures were observed in one patient. Specifically, seizures started at 3 months of age in patient 5, who was intractable to multiple antiepileptic drugs. The seizure semiology was generalized as tonic or tonic-clonic, and seizures occurred several times a day. EEG studies revealed focal epileptic discharges in the unilateral fronto-central region.

All patients had an intellectual disability, but its severity varied. Seven patients never walked independently or spoken a single word; however, they could make eye contact and social smiles, and

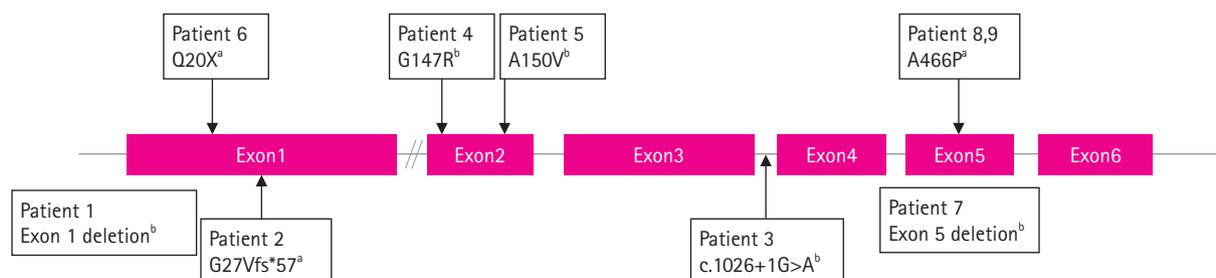


Fig. 1. Mutations of the solute carrier family 16 member 2 (*SLC16A2*) gene in this study. <sup>a</sup>Novel variant; <sup>b</sup>Previously reported variant.

recognize their parents. Patient 8 could say a few words, but was not able to make a sentence and could comprehend simple commands. Patient 9 who had the same mutation variant as patient 8 could not say any words, but could communicate with gestures and understand simple commands. Interestingly, patient 8 showed regression of language ability at the age of 7 years in both expressive and receptive language.

Cataplexy, which is a sudden loss of muscle tone triggered by emotional stress, was observed in patients 8 and 9 at the ages of 7 and 5 years, respectively. No other patients in this cohort showed cataplexy.

## 6. Brain MRI

Among the nine patients, patients 8 and 9, who had the same *SLC16A2* mutation variant, showed normal brain MRI findings on examinations performed at 1, 2, and 8 years and 1 year of age, respectively. Abnormalities on brain MRI were observed in seven patients, including delayed myelination, brain atrophy, a thin corpus callosum, and a prominent subarachnoid space in the middle cranial fossa. Delayed myelination was found in five patients. Among them, serial MRI studies were obtained in one patient and delayed myelination was persistent until the last follow-up at 4 years of age (Fig. 2). Diffuse cerebral atrophy was found in five patients, and serial MRI studies in two of them revealed progression of cerebral atrophy. A thin corpus callosum was found in four patients. In one of

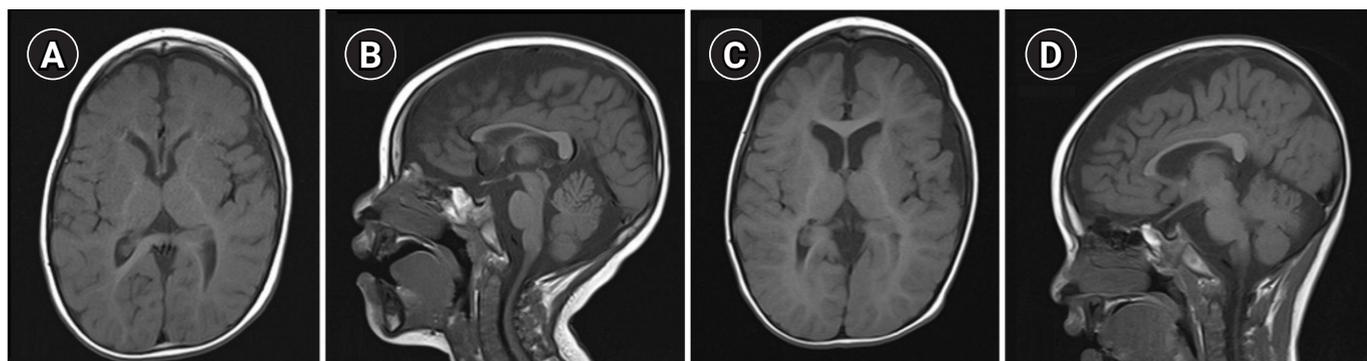
them, a thin corpus callosum was the only abnormality observed on brain MRI. A subarachnoid cyst or prominent subarachnoid space in both middle cranial fossae was found in two patients.

## 7. Thyroid hormone blood levels

The thyroid hormone profiles at patients' initial examinations are summarized in Table 2. The median age at the first test was 16 months (range, 8 to 83). TSH levels were normal in eight patients and high in patient 1. Patient 1 had low free  $T_4$  with a high TSH level; therefore, he was treated with levothyroxine replacement before the diagnosis of AHDS. Low free  $T_4$  levels were found in four patients and were normal in five patients.  $T_3$  levels were high in all nine patients at the initial test. The median  $T_3$  level at the initial test was 250 ng/dL (range, 190 to 436.5; reference, 64 to 152). In patient 5, successive thyroid hormone tests revealed normalization of  $T_3$  levels and decreased free  $T_4$  levels below the normal range at 4 and 5 years of age compared to the initial results obtained at 9 months of age. The ratio of  $T_3$  to free  $T_4$  was also calculated, and the median  $T_3$  to free  $T_4$  ratio was 420.7 (range, 231.7 to 704.0; reference, 41.2 to 217.1). The  $T_3$  to free  $T_4$  ratio was consistently high even in patient 5, whose  $T_3$  level was normalized and had a decreased free  $T_4$  level.

## 8. Laboratory abnormalities

Elevated lactic acid levels were observed in five patients. Severe lac-



**Fig. 2.** Brain magnetic resonance imaging (MRI) of patient 2 at the ages of (A, B) 22 months and (C, D) 4 years. (A, C) Axial and (B, D) sagittal T1-weighted MRI showed a thin corpus callosum, progression of cerebral atrophy, and persistent hypomyelination.

**Table 2.** Initial thyroid hormone levels

Variable	Reference	Patient 1	Patient 2	Patient 3	Patient 4	Patient 5	Patient 6	Patient 7	Patient 8	Patient 9
$T_3$ (ng/dL)	64–152	250.4	219.4	280	238	190	436.5	228	358.3	281.9
TSH ( $\mu$ IU/mL)	0.35–4.94	6.34	2.16	4.10	2.74	2.4	3.95	4.46	2.09	2.70
Free $T_4$ (ng/dL)	0.70–1.48	0.54	0.71	0.52	0.66	0.82	0.62	0.83	0.73	0.70
$T_3$ /free $T_4$	41.2–217.1	463.0	309.0	538.5	360.6	231.7	704.0	274.7	490.8	402.7

$T_3$ , triiodothyronine; TSH, thyroid-stimulating hormone;  $T_4$ , tetraiodothyronine.

tic acidosis needing alkali treatment was observed in two patients and hyperlactatemia without metabolic acidosis was observed in the remaining three patients. Transient hyperammonemia accompanied by severe lactic acidosis was observed in patient 2, which was incidentally identified from the metabolic workup and treated with sodium benzoate. Neurologic symptoms associated with hyperammonemia were not observed. After that event, the ammonia level remained in a normal range even after ceasing medication. Intermittent elevation of aspartate aminotransferase (AST), alanine aminotransferase (ALT), and creatinine kinase (CK) was noted in two patients.

### 9. Skeletal problems and other problems (urologic and ocular)

Skeletal abnormalities were found in four patients, including hip dislocation ( $n = 2$ ), bilateral coxa valga ( $n = 1$ ), macrodactyly of the first and second toes ( $n = 1$ ), and pes planovalgus ( $n = 1$ ). Cryptorchidism was present in patient 5 and testis sonography revealed unilateral gliding testis. Ocular examinations were performed in six patients, and strabismus was noted in one patient.

## Discussion

In this study, we conducted a detailed investigation of the clinical, genotypic, and laboratory characteristics of nine male patients with genetically confirmed AHDS.

After the causative gene *SLC16A2* was revealed, the pathophysiology of AHDS has been understood as resistance to thyroid hormones in the brain due to the deficiency of the thyroid hormone transporter, MCT8, which plays a key role in transporting thyroid hormones across the blood–brain barrier and into neuronal and glial cells in developing brains [13]. The MCT family, which is a member of the major facilitator superfamily (MFS), has a shared structural fold with 12 transmembrane domains (TMDs) and cytosolic N- and C-termini [14]. This transmembrane protein is thought to control the cellular influx and efflux of thyroid hormones and regulate intracellular hormone levels [4,15]. An expression analysis of MCT8 revealed that it is present in cerebral neurons, glial cells, and the cellular structures that compose the blood–brain barrier and blood–cerebrospinal fluid barrier in the fetal brain [5,16]. In addition to the brain, MCT8 is also abundant in the thyroid glands, kidneys, and liver [4].

A distinctive thyroid hormone profile is a hallmark of AHDS [2,3]. Typically, the  $T_3$  level is elevated, the free  $T_4$  level is normal to low, and the TSH level is normal to mildly elevated. Considering the diseases resulting from defects of other thyroid hormone transporters, such as OATP1C1 deficiency, which shows a normal thy-

roid hormone profile [17], abnormal serum thyroid hormone levels not only provide a clue for diagnosis, but also show the association of MCT8 and the hypothalamus-pituitary-thyroid (HPT) axis. Studies on the relevance of MCT8 to the HPT axis in *Mct8*-knock-out mice have shown that the hypothalamus and pituitary gland seem to poorly sense serum thyroid hormone concentrations, which suggests that TSH level is not a reliable indicator of the thyroid hormone status of the brain and body [18,19]. Although an elevated  $T_3$  level was thought to be a fingerprint of AHDS, normal  $T_3$  levels were reported in some cases [10,11]. In our cohort, patient 5, who had an elevated  $T_3$  level of 190 ng/dL (reference range, 64 to 152) at the age of 9 months showed normal serum  $T_3$  levels of 144.5 and 115.7 ng/dL at the ages of 4 and 5 years, respectively, with decreased free  $T_4$  levels, but the ratio of  $T_3$  to free  $T_4$  was still elevated. If AHDS is clinically suspected, it is recommended to test the patients' thyroid hormone status repeatedly and keep in mind that the  $T_3$  level can vary. An increased  $T_3$  to free  $T_4$  ratio may be a more reliable parameter for clinical suspicion of AHDS.

The types of mutations in the *SLC16A2* gene vary considerably, from single-nucleotide variants to large deletions. In our cohort, we reported three missense mutation, one nonsense mutation, one frameshift mutation, one splicing-site mutation, and two exon deletions. Several functional studies have shown that genetic mutations of *SLC16A2* cause a variable degree of functional defects in MCT8, including impairments of homodimerization, localization to the plasma membrane, and transport function [20,21]. Two previously reported missense mutations were found in patients 4 and 5, who had c.439G > A [p.G147R] and c.449C > T [p.A150V] mutations, respectively. The c.439G > A [p.G147R] mutation changes the glycine located in the second TMD to arginine. In functional studies [20], it impaired the thyroid hormone transporter activity without affecting localization to the plasma membrane. The clinical manifestation of two patients with the p.G147R mutation from the same family reported previously [18] was similar to patient 4, who had a severe phenotype with spastic paraplegia and absence of verbal output, except for seizures, which were not observed as of the age of 4 years (at his last visit). The c.449C > T [p.A150V] mutation in patient 5 changed alanine to valine in the second TMD. This has been reported previously, and all cases had the same severe phenotype as patient 5 [19,22,23]. In the functional analysis [19,21], this mutation was reported as not affecting dimer assembly [21], but impairing expression in the plasma membrane [19]. The novel 1396G > C [p.A466P] missense mutation from patients 8 and 9 changed alanine to proline in the 11th TMD and caused a relatively mild phenotype, as the patients gained the ability to walk and had normal brain MRI findings. A

functional study was not performed for these patients. In our cohort, exon deletion was found in patients 1 and 7, who showed exon 1 and exon 5 deletion, respectively; sole exon 5 deletion has not been reported previously. Both patients had a severe phenotype, with spastic paraplegia and no verbal output. Patient 3 had a c.1026+1G > A splicing site mutation, which was previously reported in one case [24]. In that case, the clinical presentation was similar, showing a severe phenotype, but the thyroid hormone profile was different because the previously reported case showed lower free T<sub>4</sub> levels than patient 3, who had normal free T<sub>4</sub> levels [24]. The novel frameshift mutation c.80delG [p.G27Vfs\*57] and non-sense mutation c.58C > T [p.Q20X] on exon 1 were observed in patients 2 and 6, respectively, who had severe phenotypes.

Axial hypotonia and spasticity were the most common features in our cohort, which corresponds well with previously reported cases [10,11]. Among several extrapyramidal phenotypes, exaggerated startles or myoclonus was a notable phenotype in this cohort, as observed in 55% (5/9) of the patients. Dystonia, which is also a commonly reported manifestation of AHDS, was also widely observed in this study.

Along with the increasing number of reported patients and genetic mutations of AHDS over the last decade, some patient groups have shown milder phenotypes than others, with mild intellectual disability or achieving ambulation [10,18,25-29]. In this cohort, patients 8 and 9, who had the same missense 1396G > C [p.A466P] mutation at the end of the 11th TMD, showed a distinctively milder phenotype compared to the other patients. Their age of onset was later than that of the others and they achieved ambulation around 5 years of age. Although both had the same variants and could be categorized as having a mild phenotype, they had phenotypic differences. At the initial neurologic examination at the age of 12 months, patient 9 showed more classical clinical features, such as hypotonia, spasticity, and hyperreflexia, whereas patient 8 showed none of these features except for mild hypotonia. During the follow-up period, these clinical features became evident in patient 8. In this patient with a mild phenotype, the initial neurologic examination was uncertain; thus, thoughtful follow-up is crucial.

To our knowledge, cataplexy, which was a notable shared phenotype of patients 8 and 9, has not previously been reported in patients with AHDS. As one of the features of narcolepsy, cataplexy is thought to be related to the deficiency of orexin, a neurotransmitter secreted by neurons in the lateral hypothalamus [30], which plays a role in the sleep-wakefulness cycle. There is a topographical similarity in that both orexin-producing neurons and MCT8-expressing cells exist in the hypothalamus [31]. Although the underlying mechanism is unclear, several studies have shown

evidence supporting the association between the HPT axis and the orexin system. In an animal study, treatment with a thyrotropin-releasing hormone analogue improved cataplexy [32] and a low level of TSH was reported in patients with orexin-deficient narcolepsy [33]. The finding of cataplexy in patients with AHDS could also support the suggested link between the two systems; thus, further research is needed.

Several phenotypes in patients with AHDS, including poor weight gain, decreased muscle bulk, and tachycardia as reported previously, were thought to be caused by peripheral thyrotoxicosis. Poor weight gain was also a common phenotype in this cohort. Skeletal muscle is one of the major target organs for thyroid hormones, which regulate homeostasis. The regeneration of skeletal muscle and excessive T<sub>3</sub> levels in patients with AHDS changes the expression pattern of numerous proteins toward a catabolic status, which results in muscle wasting [34]. Elevation of lactic acid was observed in about half of the patients, and one-third of them experienced metabolic acidosis. Along with lactic acid, the increased levels of AST, ALT, CK, and ammonia were thought to reflect the catabolic status of skeletal muscle [8]. These laboratory abnormalities can sometimes lead to the misdiagnosis of mitochondrial or muscle disease; for example, previous reports of brain MRI findings in patients with AHDS [6] showed that delayed myelination or hypomyelination was a major accompanying feature with or without cerebral atrophy. In some previous research, normalization of myelination in the successive follow-up of brain MRI was reported [6,7]; however, this was not observed in our case. The observation of severe hypomyelination on brain imaging can lead to a misdiagnosis of other neurodegenerative diseases that are categorized into leukodystrophy, such as lysosomal storage disease or mitochondrial disease. Normal brain MRI findings were also observed in two patients with milder phenotypes, which makes AHDS difficult to diagnose.

Due to the specific pathophysiology and significant burden of AHDS, various efforts have been made to obtain an effective treatment [35]. Recently, a second-phase clinical trial of treatment with triiodothyroacetic acid, which is a thyroid hormone analogue, showed clinical improvement of peripheral thyrotoxicosis and the possibility of improving patients' neurologic status if treated at a young age [36]. Therefore, an early diagnosis is becoming more important.

Herein, we report nine patients who were genetically confirmed with AHDS with mutations of *SLC16A2*, which encodes MCT8, a thyroid hormone transporter. Phenotypic and genetic variability is a notable feature of the disease, which makes an early diagnosis difficult. Understanding its broad clinical features, which reflect the opposite patterns of thyroid hormone status in the brain and pe-

riphery, with a variable serum thyroid profile and MRI findings will help to stimulate clinical suspicion and guide early confirmatory genetic tests. Importantly, a thyroid function test including T<sub>3</sub> levels should be considered when encountering male patients with severe developmental delay and spasticity.

## Conflicts of interest

Jieun Choi is an associate editor, Ki Joong Kim and Jong-Hee Chae are the editorial board members of the journal, but They was not involved in the peer reviewer selection, evaluation, or decision process of this article. No other potential conflicts of interest relevant to this article were reported.

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## References

- Allan W, Herndon CN, Dudley FC, Allan W, Nash Herndon C, Dudley EC, et al. Some examples of the inheritance of mental deficiency: apparently sex-linked idiocy and microcephaly. *Am J Ment Defic* 1944;48:325-34.
- Dumitrescu AM, Liao XH, Best TB, Brockmann K, Refetoff S. A novel syndrome combining thyroid and neurological abnormalities is associated with mutations in a monocarboxylate transporter gene. *Am J Hum Genet* 2004;74:168-75.
- Friesema EC, Grueters A, Biebermann H, Krude H, von Moers A, Reeser M, et al. Association between mutations in a thyroid hormone transporter and severe X-linked psychomotor retardation. *Lancet* 2004;364:1435-7.
- Friesema EC, Ganguly S, Abdalla A, Manning Fox JE, Halestrap AP, Visser TJ. Identification of monocarboxylate transporter 8 as a specific thyroid hormone transporter. *J Biol Chem* 2003;278:40128-35.
- Roberts LM, Woodford K, Zhou M, Black DS, Haggerty JE, Tate EH, et al. Expression of the thyroid hormone transporters monocarboxylate transporter-8 (SLC16A2) and organic ion transporter-14 (SLCO1C1) at the blood-brain barrier. *Endocrinology* 2008;149:6251-61.
- Vancamp P, Demeneix BA, Remaud S. Monocarboxylate transporter 8 deficiency: delayed or permanent hypomyelination? *Front Endocrinol (Lausanne)* 2020;11:283.
- Azzolini S, Nosadini M, Balzarin M, Sartori S, Suppiej A, Mardari R, et al. Delayed myelination is not a constant feature of Allan-Herndon-Dudley syndrome: report of a new case and review of the literature. *Brain Dev* 2014;36:716-20.
- Herzovich V, Vaiani E, Marino R, Dratler G, Lazzati JM, Tilitzky S, et al. Unexpected peripheral markers of thyroid function in a patient with a novel mutation of the MCT8 thyroid hormone transporter gene. *Horm Res* 2007;67:1-6.
- Dumitrescu AM, Liao XH, Weiss RE, Millen K, Refetoff S. Tissue-specific thyroid hormone deprivation and excess in monocarboxylate transporter (mct) 8-deficient mice. *Endocrinology* 2006;147:4036-43.
- Remerand G, Boespflug-Tanguy O, Tonduti D, Touraine R, Rodriguez D, Curie A, et al. Expanding the phenotypic spectrum of Allan-Herndon-Dudley syndrome in patients with SLC16A2 mutations. *Dev Med Child Neurol* 2019;61:1439-47.
- Groeneweg S, van Geest FS, Abaci A, Alcantud A, Ambegaonkar GP, Armour CM, et al. Disease characteristics of MCT8 deficiency: an international, retrospective, multicentre cohort study. *Lancet Diabetes Endocrinol* 2020;8:594-605.
- Jansen J, Friesema EC, Kester MH, Schwartz CE, Visser TJ. Genotype-phenotype relationship in patients with mutations in thyroid hormone transporter MCT8. *Endocrinology* 2008;149:2184-90.
- Heuer H, Maier MK, Iden S, Mittag J, Friesema EC, Visser TJ, et al. The monocarboxylate transporter 8 linked to human psychomotor retardation is highly expressed in thyroid hormone-sensitive neuron populations. *Endocrinology* 2005;146:1701-6.
- Lafreniere RG, Carrel L, Willard HF. A novel transmembrane transporter encoded by the XPCT gene in Xq13.2. *Hum Mol Genet* 1994;3:1133-9.
- Friesema EC, Jansen J, Jachtenberg JW, Visser WE, Kester MH, Visser TJ. Effective cellular uptake and efflux of thyroid hormone by human monocarboxylate transporter 10. *Mol Endo-*

- crinol 2008;22:1357-69.
16. Lopez-Espindola D, Garcia-Aldea A, Gomez de la Riva I, Rodriguez-Garcia AM, Salvatore D, Visser TJ, et al. Thyroid hormone availability in the human fetal brain: novel entry pathways and role of radial glia. *Brain Struct Funct* 2019;224:2103-19.
  17. Stromme P, Groeneweg S, Lima de Souza EC, Zevenbergen C, Torgersbraten A, Holmgren A, et al. Mutated thyroid hormone transporter OATP1C1 associates with severe brain hypometabolism and juvenile neurodegeneration. *Thyroid* 2018;28:1406-15.
  18. Vaur-Barriere C, Deville M, Sarret C, Giraud G, Des Portes V, Prats-Vinas JM, et al. Pelizaeus-Merzbacher-Like disease presentation of MCT8 mutated male subjects. *Ann Neurol* 2009;65:114-8.
  19. Biebermann H, Ambrugger P, Tarnow P, von Moers A, Schweizer U, Grueters A. Extended clinical phenotype, endocrine investigations and functional studies of a loss-of-function mutation A150V in the thyroid hormone specific transporter MCT8. *Eur J Endocrinol* 2005;153:359-66.
  20. Kersseboom S, Kremers GJ, Friesema EC, Visser WE, Klootwijk W, Peeters RP, et al. Mutations in MCT8 in patients with Allan-Herndon-Dudley-syndrome affecting its cellular distribution. *Mol Endocrinol* 2013;27:801-13.
  21. Fischer J, Kleinau G, Muller A, Kuhnen P, Zwanziger D, Kinne A, et al. Modulation of monocarboxylate transporter 8 oligomerization by specific pathogenic mutations. *J Mol Endocrinol* 2015;54:39-50.
  22. Choi JH, Cho JH, Kim JH, Yoo EG, Kim GH, Yoo HW. Variable clinical characteristics and molecular spectrum of patients with syndromes of reduced sensitivity to thyroid hormone: genetic defects in the THRB and SLC16A2 genes. *Horm Res Paediatr* 2018;90:283-90.
  23. Jansen J, Friesema EC, Kester MH, Milici C, Reeser M, Gruters A, et al. Functional analysis of monocarboxylate transporter 8 mutations identified in patients with X-linked psychomotor retardation and elevated serum triiodothyronine. *J Clin Endocrinol Metab* 2007;92:2378-81.
  24. Wang A, Xi J, Yuan F, Wang Y, Wang S, Wang C, et al. Generation of an induced pluripotent stem cell line (SHCDNi003-A) from a one-year-old Chinese Han infant with Allan-Herndon-Dudley syndrome. *Stem Cell Res* 2020;46:101872.
  25. Schwartz CE, May MM, Carpenter NJ, Rogers RC, Martin J, Bialer MG, et al. Allan-Herndon-Dudley syndrome and the monocarboxylate transporter 8 (MCT8) gene. *Am J Hum Genet* 2005;77:41-53.
  26. Visser WE, Jansen J, Friesema EC, Kester MH, Mancilla E, Lundgren J, et al. Novel pathogenic mechanism suggested by ex vivo analysis of MCT8 (SLC16A2) mutations. *Hum Mutat* 2009;30:29-38.
  27. Novara F, Groeneweg S, Freri E, Estienne M, Reho P, Matricardi S, et al. Clinical and molecular characteristics of SLC16A2 (MCT8) mutations in three families with the Allan-Herndon-Dudley syndrome. *Hum Mutat* 2017;38:260-4.
  28. Braun D, Schweizer U. The chemical chaperone phenylbutyrate rescues MCT8 mutations associated with milder phenotypes in patients with Allan-Herndon-Dudley syndrome. *Endocrinology* 2017;158:678-91.
  29. Masnada S, Groeneweg S, Saletti V, Chiapparini L, Castellotti B, Salsano E, et al. Novel mutations in SLC16A2 associated with a less severe phenotype of MCT8 deficiency. *Metab Brain Dis* 2019;34:1565-75.
  30. Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, et al. Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci* 1998;18:9996-10015.
  31. Alkemade A, Friesema EC, Unmehopa UA, Fabriek BO, Kuiper GG, Leonard JL, et al. Neuroanatomical pathways for thyroid hormone feedback in the human hypothalamus. *J Clin Endocrinol Metab* 2005;90:4322-34.
  32. Riehl J, Honda K, Kwan M, Hong J, Mignot E, Nishino S. Chronic oral administration of CG-3703, a thyrotropin releasing hormone analog, increases wake and decreases cataplexy in canine narcolepsy. *Neuropsychopharmacology* 2000;23:34-45.
  33. Kok SW, Roelfsema F, Overeem S, Lammers GJ, Frolich M, Meinders AE, et al. Altered setting of the pituitary-thyroid ensemble in hypocretin-deficient narcoleptic men. *Am J Physiol Endocrinol Metab* 2005;288:E892-9.
  34. Mayerl S, Schmidt M, Doycheva D, Darras VM, Huttner SS, Boelen A, et al. Thyroid hormone transporters MCT8 and OATP1C1 control skeletal muscle regeneration. *Stem Cell Reports* 2018;10:1959-74.
  35. Grijota-Martinez C, Barez-Lopez S, Gomez-Andres D, Guadano-Ferraz A. MCT8 deficiency: the road to therapies for a rare disease. *Front Neurosci* 2020;14:380.
  36. Groeneweg S, Peeters RP, Moran C, Stoupa A, Auriol F, Tonduti D, et al. Effectiveness and safety of the tri-iodothyronine analogue Triac in children and adults with MCT8 deficiency: an international, single-arm, open-label, phase 2 trial. *Lancet Diabetes Endocrinol* 2019;7:695-706.